Europe (8), resident populations in these countries have been exposed to these virus lineages more frequently than populations in Asia, and therefore may have acquired a greater degree of preexisting cross-reactive immunity to pandemic (H1N1) 2009 virus. A recent review of human swine influenza infections suggests that they may not be uncommon (9), although the true incidence of human infections with swine influenza is unknown because of paucity of swine influenza surveillance data worldwide (8).

In conclusion, partial cross-immunity and cell-mediated immunity may be present but not detected by HI or MN assays. Thus, results of standard serologic assays may not be providing all relevant data (10).

Testing by the Melbourne World Health Organization Collaborating Centre for Reference and Research on Influenza was supported by the Australian Government Department of Health and Ageing.

Julian W. Tang, Paul A. Tambyah, Annelies Wilder-Smith, Kim-Yoong Puong, Robert Shaw, Ian G. Barr, and Kwai-Peng Chan

Author affiliations: National University Hospital, Singapore (J.W. Tang); National University of Singapore, Singapore (P.A. Tambyah, A. Wilder-Smith); Singapore General Hospital, Singapore (K.-Y. Puong, K.-P. Chan); and World Health Organization Laboratory for Influenza Reference and Research, Melbourne, Victoria, Australia (R. Shaw, I. Barr)

DOI: 10.3201/eid1605.091678

References

 World Health Organization. WHO manual on animal influenza diagnosis and surveillance. Geneva: The Organization. Document WHO/CDS/CSR/NCS /2002.5); 2002. p. 28–39.

- World Health Organization. WHO manual on animal influenza diagnosis and surveillance. Geneva: The Organization. Document WHO/CDS/CSR/NCS /2002.5); 2002. p. 48–54.
- Chen H, Wang Y, Liu W, Zhang J, Dong B, Fan X, et al. Serologic survey of pandemic (H1N1) 2009 virus, Guanxi Province, China. Emerg Infect Dis. 2009;15:1849–50.
- Itoh Y, Shinya K, Kiso M, Watanabe T, Sakoda Y, Hatta M, et al. In vitro and in vivo characterization of new swineorigin H1N1 influenza viruses. Nature. 2009:460:1021–5
- Hancock K, Veguilla V, Lu X, Zhong W, Butler EN, Sun H, et al. Cross-reactive antibody responses to the 2009 pandemic H1N1 influenza virus. N Engl J Med. 2009;361:1945–52. DOI: 10.1056/NEJ-Moa0906453
- Macroepidemiology of Influenza Vaccination (MIV) Study Group. The macroepidemiology of influenza vaccination in 56 countries, 1997–2003. Vaccine. 2005;23:5133–43. DOI: 10.1016/j. vaccine.2005.06.010
- Miller E, Hoschler K, Hardelid P, Standford E, Andrews N, Zambon M. Incidence of 2009 pandemic influenza A H1N1 infection in England: a cross-sectional serological study. Lancet. 2010; Jan 20. [Epub ahead of print].
- Peiris JS, Poon LL, Guan Y. Emergence of a novel swine-origin influenza A virus (S-OIV) H1N1 virus in humans. J Clin Virol. 2009;45:169–73. DOI: 10.1016/j. jcv.2009.06.006
- Myers KP, Olsen CW, Gray GC. Cases of swine influenza in humans: a review of the literature. Clin Infect Dis. 2007;44:1084–8. DOI: 10.1086/512813
- Xing Z, Cardona CJ. Preexisting immunity to pandemic (H1N1) 2009. Emerg Infect Dis. 2009;15:1847–9.

Address for correspondence: Julian W. Tang, Division of Microbiology/Molecular Diagnostic Centre, Department of Laboratory Medicine, National University Hospital, 5 Lower Kent Ridge Rd, 119074 Singapore; email: jwtang49@hotmail.com



Molecular Epidemiology of Japanese Encephalitis Virus, Taiwan

To the Editor: Japanese encephalitis virus (JEV) is a mosquito-borne member of the family Flaviviridae and the genus Flavivirus. JEV is a major cause of viral encephalitis in Asia. Phylogenetic analysis of the envelope (E) gene sequences has shown that JEV strains can be clustered into 5 distinct genotypes (1). Among them, genotype III (GIII) has had the widest geographic distribution in countries in Asia, including Japan, South Korea, People's Republic of China, Taiwan, Vietnam, the Philippines, and India (2). Before 1990, GIII had been the major epidemic JEV type in these areas. However, the introduction of JEV genotype I (GI) has been reported in Japan, Vietnam, South Korea, Thailand, and China in the past decade (3–6). Nabeshima et al. recently reported surveillance results that provided substantial evidence of frequent introductions of JEV GI into Japan from Southeast Asia and continental eastern Asia (7). Because all current vaccines are derived from JEV GIII strains, the effectiveness of vaccination in inducing protective neutralizing antibodies against various genotype strains needs to be carefully evaluated, taking into account genotype shift in these countries.

Japanese encephalitis is endemic in Taiwan. Reports on the molecular epidemiology of JEV in Taiwan are scarce. Jan et al. (8) reported the genetic variation of 47 JEV isolates from Taiwan before 1994. Phylogenetic analysis showed that all Taiwanese isolates were GIII, and they were classified into 3 clusters.

To understand the genetic variation of JEV strains currently circulating in Taiwan, we conducted a surveillance program in the following areas: northern (Taipei, Taoyuan, and Yilan counties and Taipei City), central (Taichung and Changhua counties), southern (Tainan and Kaohsiung counties), and eastern (Hualien County) during 2005-2008. Real-time reverse transcription-PCR (RT-PCR) was used to screen JEV in mosquito pools, pig serum specimens, and human cerebrospinal fluid as described (9). Mosquitoes were pooled by species, location, and collection date in groups of 30-50 mosquitoes. Mosquito pools were homogenized and clarified by centrifugation, and the supernatants were sterilized by filtration and removed for real-time RT-PCR and virus isolation.

We used 3 sets of primers for realtime RT-PCR: flavivirus-specific (FL-F1: 5'-GCCATATGG TACATGTG-GCTGGGAGC-3'; FL-R3: 5'-GTKA TTCTTGTGTCCCAWCCGGCTGT GTCATC-3'; FL-R4: 5'-GTGATGCG RGTGTCCCAGCCRGCKGTGT CATC-3'), JEV-specific (10) (JE3F1: 5'-CCCTCAGAACCGTCTCGGAA-3' and JE3R1: 5'-CTATTCCCAGGTG TCAATATGCTGT-3'), and JEV GIIIspecific (E12F: 5'-CTGGGAATGG GCAATCGTG-3' and E325R: 5'-TGTCAATGCTTCCCTTCCC-3'). Samples with positive results by RT-PCR were subjected to virus isolation by using a mosquito C6/36 cell line. A total of 47 JEV isolates were obtained: 38 from mosquitoes, 8 from pig serum samples, and 1 from human cerebrospinal fluid.

Viral RNA was extracted from JEV-infected culture medium, and RT-PCR and DNA sequencing were performed to determine the complete E gene sequences of JEV isolates. Multiple sequence alignment and phylogenetic analysis were conducted by using CLUSTALW software (www. ebi.ac.uk/Tools/clustalw2/index.html) and MEGA version 4 (www.megasofteware.net). The phylogenetic tree was constructed by the neighbor-joining method and the maximum composite likelihood model.

The Figure shows the phylogenetic tree derived from 67 samples of

E gene sequences, including 28 representative new sequences in this study (GenBank accession nos. GQ260608–GQ260635), 10 sequences of Taiwanese strains isolated before 2002, and

29 sequences from GenBank. The results show that isolates from Taiwan comprised 2 genotypes, GIII and GI. All of the JEV isolates from Taiwan obtained during 2005–2008, except

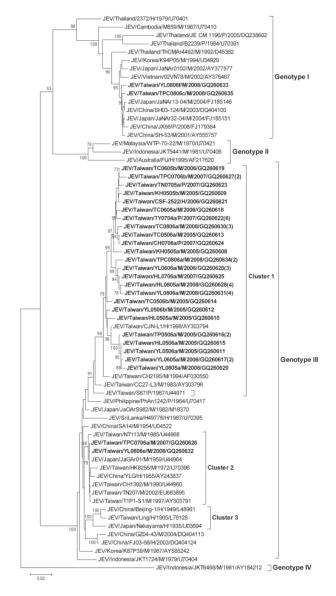


Figure. Phylogenetic tree showing the genetic relationship among Japanese encephalitis virus (JEV) isolates. The tree was constructed on the basis of complete envelope (E) nucleotide sequences of JEV strains. Sequences obtained in this study are indicated in **boldface**. Genotypes are indicated on the right. Viruses were identified by using the nomenclature of virus/country/strain/source/year of isolation/GenBank accession number. Numbers in parentheses indicate the number of isolates that showed 100% nucleotide homology. Isolates with the same sequences were collected at the same time from the same location in this study. Analysis was performed by using MEGA 4 software (www. megasoftware.net) and neighbor-joining (maximum composite likelihood) methods. Bootstrap support values >75 are shown (1,000 replicates). CH, Changhua County; HL, Hualien County; KH, Kaohsiung County; TC, Taichung County; TN, Tainan County; TP, Taipei County; TPC, Taipei City; TY, Taoyuan County; YL, Yilan County; M, mosquito pool; p, pig serum; H, human sample. Scale bar indicates nucleotide substitutions per site.

2 strains (TPC0806c/M/2008 and YL0806f/M/2008), belonged to GIII and formed into 2 clusters.

Cluster 1 contains most new isolates prevalent in different areas of Taiwan. Although cluster 1 isolates are closely related to other JEV strains from Asia, these isolates, together with previously published JEV sequences from Taiwan, form a distinct lineage and may have been continuously evolving and locally adapting in Taiwan. Cluster 2 contains only 2 new isolates, TPC0706a/M/2007 and YL0806e/M/2008, which were isolated from the Culex tritaeniorhynchus mosquito pools in Kuantu Nature Park, Taipei City, and from a pig farm in Wujie Township, Yilan County, respectively.

Notably, the 2 GI strains, TPC0806c/M/2008 and YL0806f/ M/2008, were isolated from the same areas as the GIII cluster 2 strains. These areas are adjacent to the wetlands, which are stopover sites for migratory birds. These 2 GI strains are most closely related to the strains of the subcluster II JEV strains reported by Nabeshima et al. (7). The TPC0806c/M/2008 GI strain is most closely related to Japan/JaNAr13-04/M/2004 and China/SH03-124-/M/2003 strains (99.5% and 99.4% identities, respectively), and the YL0806f/M/2008 GI strain is most closely related to Japan/JaNAr13-04-/M/2004 and China/JX66/P/2008 strains (99.3% and 99.3% identities, respectively). Therefore, JEV GI strains from Taiwan were likely introduced by water birds migrating back and forth along the Asia-Australasia flyway, which passes through many countries, including Indonesia, Malaysia, Australia, the Philippines, Taiwan, China, and Japan (3).

Our results clearly showed that JEV GIII strains remain the most dominant population circulating in Taiwan, although 2 JEV GI strains were isolated from wetland areas in northern Taiwan in 2008. Further stud-

ies are needed to continuously monitor the changing epidemiologic pattern of JEV strains endemic in Taiwan and newly introduced viruses.

This study was supported in part by grant 98-0324-01-F-20 from the National Research Program for Genome Medicine, by grant DOH97-DC-2002 from Centers for Disease Control, Department of Health, Taipei, Taiwan, Republic of China, and by a grant from Ministry of Health, Labor and Welfare of Japan through the National Institute of Infectious Diseases (Tokyo).

Jyh-Hsiung Huang,
Ting-Hsiang Lin, Hwa-Jen Teng,
Chien-Ling Su, Kun-Hsien Tsai,
Liang-Chen Lu, Cheo Lin,
Cheng-Fen Yang, Shu-Fen Chang,
Tsai-Ling Liao, Sheng-Kai Yu,
Chia-Hsin Cheng,
Mei-Chun Chang, Huai-Chin Hu,
and Pei-Yun Shu

Author affiliations: Centers for Disease Control, Taipei, Taiwan (J.-H. Huang, T.-H. Lin, H.-J. Teng, C.-L. Su, L.-C. Lu, C. Lin, C.-F. Yang, S.-F. Chang, T.-L. Liao, S.-K. Yu, C.-H. Cheng, M.-C. Chang, H.-C. Hu, P.-Y. Shu); and National Taiwan University, Taipei (K.-H. Tsai)

DOI: 10.3201/eid1605.091055

References

- Solomon T, Ni H, Beasley DW, Ekkelenkamp M, Cardosa MJ, Barrett AD. Origin and evolution of Japanese encephalitis virus in southeast Asia. J Virol. 2003;77:3091–8. DOI: 10.1128/ JVI.77.5.3091-3098.2003
- Mackenzie JS, Gubler DJ, Petersen LR. Emerging flaviviruses: the spread and resurgence of Japanese encephalitis, West Nile and dengue viruses. Nat Med. 2004;10(Suppl):S98–109. DOI: 10.1038/ nm1144
- Ma SP, Yoshida Y, Makino Y, Tadano M, Ono T, Ogawa M. A major genotype of Japanese encephalitis virus currently circulating in Japan. Am J Trop Med Hyg. 2003:69:151–4.
- Nga PT, del Carmen Parquet M, Cuong VD, Ma SP, Hasebe F, Inoue S, et al. Shift in Japanese encephalitis virus (JEV) genotype circulating in northern Viet-

- nam: implications for frequent introductions of JEV from Southeast Asia to East Asia. J Gen Virol. 2004;85:1625–31. DOI: 10.1099/vir.0.79797-0
- Nitatpattana N, Dubot-Pérès A, Gouilh MA, Souris M, Barbazan P, Yoksan S, et al. Change in Japanese encephalitis virus distribution, Thailand. Emerg Infect Dis. 2008;14:1762–5. DOI: 10.3201/ eid1411.080542
- Wang HY, Takasaki T, Fu SH, Sun XH, Zhang HL, Wang ZX, et al. Molecular epidemiological analysis of Japanese encephalitis virus in China. J Gen Virol. 2007;88:885–94. DOI: 10.1099/ vir.0.82185-0
- Nabeshima T, Loan HT, Inoue S, Sumiyoshi M, Haruta Y, Nga PT, et al. Evidence of frequent introductions of Japanese encephalitis virus from south-east Asia and continental east Asia to Japan. J Gen Virol. 2009;90:827–32. DOI: 10.1099/ vir.0.007617-0
- Jan LR, Yueh YY, Wu YC, Horng CB, Wang GR. Genetic variation of Japanese encephalitis virus in Taiwan. Am J Trop Med Hyg. 2000;62:446–52.
- Shu PY, Chang SF, Kuo YC, Yueh YY, Chien LJ, Sue CL, et al. Development of group- and serotype-specific one-step SYBR green I-based real-time reverse transcription-PCR assay for dengue virus. J Clin Microbiol. 2003;41:2408–16. DOI: 10.1128/JCM.41.6.2408-2416.2003
- Jeong HS, Shin JH, Park YN, Choi JY, Kim YL, Kim BG, et al. Development of real-time RT-PCR for evaluation of JEV clearance during purification of HPV type 16 L1 virus-like particles. Biologicals. 2003;31:223-9. DOI: 10.1016/S1045-1056(03)00064-2

Address for correspondence: Pei-Yun Shu, Research and Diagnostic Center, Centers for Disease Control, Department of Health, 161, Kun-Yang St, Taipei, Taiwan; email: pyshu@cdc.gov.tw

