

- S segment from Jujutiba hantavirus: identification of two distinct lineages in *Oligoryzomys nigripes*. *Infect Genet Evol.* 2013;18:262–8. <http://dx.doi.org/10.1016/j.meegid.2013.05.027>
8. Ronquist F, Huelsenbeck JP. MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics.* 2003;19:1572–4. <http://dx.doi.org/10.1093/bioinformatics/btg180>
 9. dos Santos MC, De Lacerda MVG, Benedetti SM, Albuquerque BC, De Aguiar Filho AABV, Elkhoury MDR, et al. Human hantavirus infection, Brazilian Amazon. *Emerg Infect Dis.* 2006;12:1165–7. <http://dx.doi.org/10.3201/eid1207.060074>
 10. Musser GG, Carleton MD. Superfamily Muroidea. In: Wilson DE, Reeder DM, editors. *Mammal species of the world: a taxonomic and geographic reference*. 3rd ed. Baltimore: Johns Hopkins University Press; 2005. p. 849–1531.

Address for correspondence: Renata C. de Oliveira, Pavilhão Hélio e Peggy Pereira, Sala B115, Instituto Oswaldo Cruz, FIOCRUZ, Avenida Brasil 4365, Manguinhos, 21040-900, Rio de Janeiro/RJ, Brazil; email: reoliveira@ioc.fiocruz.br

Reemergence of *Brucella melitensis* in Wildlife, France

To the Editor: Brucellosis is a worldwide zoonosis caused by *Brucella* spp. France has been free of bovine, ovine, and caprine brucellosis (caused by *B. abortus* or *B. melitensis*) since 2003 (1). In early 2012, an outbreak of bovine and human brucellosis caused by *B. melitensis* biovar 3 (*Bme13*) occurred in a French Alp massif (mountainous region), where the last reported outbreak occurred in 1999 (online Technical Appendix Figure, <http://wwwnc.cdc.gov/EID/article/20/9/13-1517-Techapp1.pdf>) (2). This outbreak suggested the

persistence or reemergence of *Brucella* spp. in livestock.

An extensive investigation was conducted that involved 40 animal herds with direct links to the outbreak. Six months later, blood samples from each adult animal in any herd (12,116 animals in 205 herds) that grazed during the summer of 2012 in the massif underwent serologic analysis. However, no other case was identified in this population (online Technical Appendix Table 1). Therefore, a potential wildlife source was investigated.

Wild ruminants in the study area were the following species: hunted red deer (*Cervus elaphus*), roe deer (*Capreolus capreolus*), chamois (*Rupicapra rupicapra*), and protected Alpine ibex (*Capra ibex*). Although *B. abortus* and *B. suis* have been reported in numerous wildlife species (3), *B. melitensis* has rarely been isolated from wildlife, and only sporadic cases of infection have been reported in Europe, in chamois and Alpine ibex in the Alps (4,5) and in Iberian ibex (*Capra pyrenaica hispanica*) in the Pyrenees (6). These cases were considered to be caused by spillover from domestic ruminants, which suggests that these wild species are unable to sustain the infection (3).

We conducted our investigation during the fall–winter of 2012–2013 in the entire massif where the outbreak occurred. Blood, lung, spleen, and testes or uterus samples were obtained from all hunted animals. French Authorities authorized the killing of 12 seropositive or diseased Alpine ibex with clinical signs of brucellosis (i.e., arthritis or orchitis) among 30 captured animals.

All serum samples were tested according to requirements of the World Organisation for Animal Health for diagnosis of brucellosis in small ruminants by using by the Rose Bengal test (RBT) and the complement fixation test (CFT) (7), and by indirect ELISA (IDEXX, Montpellier, France) and competitive ELISA

(cELISA; Ingenasa, Madrid, Spain). When blood samples were unsuitable for RBT or CFT or were missing, a lung extract was tested by only the 2 ELISAs. Culture was only performed on samples from seropositive animals (online Technical Appendix Table 1) (8). If bacteriologic results were negative, a *Brucella* genus–specific real-time PCR was also used (9).

A total of 129 hunted ruminants (55 chamois, 30 red deer, 44 roe deer) were tested. No clinical signs were observed, except for arthritis in the knee of 1 chamois. All ruminants were seronegative except for the chamois, which showed positive results in the RBT, CFT, and cELISA, and 1 red deer, which showed a weakly positive result in the cELISA, but negative results by culture and real-time PCR. *Bme13* was isolated from the chamois (online Appendix Table 1).

Among 289 Alpine ibex observed in the massif, 24 were killed (22 randomly sampled animals that showed 2 diagnostic lesions at necropsy [arthritis in the knee and mammary abscesses] and 2 diseased animals [arthritis in the knee and orchitis]), and samples from these animals were subjected to serologic analysis. Ten Alpine ibex (including the 2 diseased animals) showed positive results in the RBT, CFT, and both ELISAs, and 2 showed positive results only for both ELISAs. Thus, the prevalence of *B. melitensis* in randomly captured animals was 45% (10/22; 95% CI 24.6%–66.3%) (online Technical Appendix Table 1).

Bme13 was isolated from 5 of 11 seropositive Alpine ibex (1 Alpine ibex was killed in an avalanche). Three seropositive but culture-negative ibex showed positive results by PCR (online Technical Appendix Table 2). Multilocus variable number tandem repeat analysis showed similarity among all strains isolated in this study and strains isolated from local domestic outbreaks >13 years ago (10).

Although persistence of *B. melitensis* in wild ruminants has not been reported, and these animals are considered an epidemiologic dead-end reservoir (3), the unexpected prevalence observed ($\approx 50\%$) suggests that Alpine ibex could be the source of bovine brucellosis reemergence in the study area in France. Strict surveillance policies have prevented infection of domestic livestock with *B. melitensis* in the study area since 1999. However, cohabitation of domestic and wild ruminants on pastures during the summer is rare but possible. Clinical signs and lesions observed in chamois and Alpine ibex are consistent with those reported for chamois and Alpine ibex with brucellosis (4,5). However, positive cultures were obtained from conventional target organs (knee, testes, and lymph nodes) but also from urogenital fluids, which indicates the potential for excretion of the organism.

The fact that births occur during periods and in places where female Alpine ibex are not in close contact with other wild/domestic species (because of higher altitude or rocky peaks) could explain the low transmission rate of *B. melitensis* to these animals. It also suggests that the venereal route might contribute to transmission within Alpine ibex during the mating season in winter. This report demonstrates the need for maintaining an active/reactive surveillance system for livestock and wildlife in brucellosis-free regions.

Acknowledgments

We thank Gilles Le Carrou and Yannick Corde for providing technical support; Gabriela Vecchio for providing logistic expertise; Didier Calavas for providing epidemiologic information; and Manuel Thuault, Dominique Gauthier, the Departmental Hunting Association of Haute-Savoie, local laboratories of Savoie and Haute-Savoie; and local and central veterinary services for providing efficient collaboration in the field.

**Bruno Garin-Bastuji,¹
Jean Hars,¹ Antoine Drapeau,
Moulay-Ali Cherfa,
Yvette Game,
Jean-Marie Le Horgne,
S  verine Rautureau,
Eric Maucci,
Jean-Jacques Pasquier,
Maryne Jay, and Virginie Mick**

Author affiliations: French Agency for Food, Environmental, and Occupational Health and Safety, Maisons-Alfort, France (B. Garin-Bastuji, A. Drapeau, M.-A. Cherfa, M. Jay, V. Mick); French Hunting and Wildlife Agency, Gi eres, France (J. Hars); Departmental Veterinary Laboratory of Savoie, Chamb ery, France (Y. Game); Departmental Veterinary Services of Haute-Savoie, Chamb ery (J.-M. Le Horgne); French Ministry of Agriculture, Agro-Food Industry, and Forest, Paris, France (S. Rautureau); Departmental Veterinary Laboratory of Haute-Savoie, Seynod, France (E. Maucci); and Departmental Hunters Association of Haute-Savoie, Villy-le-Pelloux, France (J.-J. Pasquier)

DOI: <http://dx.doi.org/10.3201/eid2009.131517>

References

- Rautureau S, Garin-Bastuji B, Dufour B. No brucellosis outbreak detected in sheep and goats in France in 2011 [in French]. *Bulletin  pid miologique Sant  Animale et Alimentation* 2012;54:16–9.
- Mailles A, Rautureau S, Le Horgne JM, Poinet-Leroux B, d'Arnoux C, Denneriere G, et al. Re-emergence of brucellosis in cattle in France and risk for human health. *Euro Surveill.* 2012;17:pii:20227.
- Godfroid J, Garin-Bastuji B, Saegerman C, Blasco JM. Brucellosis in terrestrial wildlife. *Rev Sci Tech.* 2013;32:27–42.
- Garin-Bastuji B, Oudar J, Richard Y, Gastellu J. Isolation of *B. melitensis* biovar 3 from a chamois (*Rupicapra rupicapra*) in the Southern French Alps. *J Wildl Dis.* 1990;26:116–8. <http://dx.doi.org/10.7589/0090-3558-26.1.116>
- Ferroglio E, Tolari F, Bollo E, Bassano B. Isolation of *Brucella melitensis* from alpine ibex. *J Wildl Dis.* 1998;34:400–2. <http://dx.doi.org/10.7589/0090-3558-34.2.400>
- Mu oz PM, Boadella M, Arnal M, de Miguel MJ, Revilla M, Martinez D, et al. Spatial distribution and risk factors of brucellosis in Iberian wild ungulates. *BMC Infect Dis.* 2010;10:46. <http://dx.doi.org/10.1186/1471-2334-10-46>
- World Organization for Animal Health (OIE). Chapter 2.4.3. Bovine brucellosis. In: *Manual of diagnostic tests and vaccines for terrestrial animals*. Paris: OIE; 2009 [cited 2014 May 1]. <http://www.oie.int>.
- Alton GG, Jones LM, Angus RD, Verger JM. *Techniques for the brucellosis laboratory*. Paris: INRA Publications; 1988.
- Bounaadja L, Albert D, Chenais B, Henault S, Zygmunt MS, Poliak S, et al. Real-time PCR for identification of *Brucella* spp.: a comparative study of IS711, *bcsp31* and *per* target genes. *Vet Microbiol.* 2009;137:156–64. <http://dx.doi.org/10.1016/j.vetmic.2008.12.023>
- Mick V, Le Carrou G, Corde Y, Game Y, Jay M, Garin-Bastuji B. *Brucella melitensis* in France: persistence in wildlife and probable spillover from alpine ibex to domestic animals. *PLoS ONE.* 2014;9:e94168. <http://dx.doi.org/10.1371/journal.pone.0094168>

Address for correspondence: Bruno Garin-Bastuji, Laboratoire de Sant  Animale, Unit  Zoonoses Bact riennes, Agence Nationale de S curit  Sanitaire de l'Alimentation, de l'Environnement et du Travail, 23 Avenue du G n ral-de-Gaulle, 94706 Maisons-Alfort Cedex, France; e-mail: bruno.garin-bastuji@anses.fr

***Clostridium tetani* Osteitis without Tetanus**

To the Editor: Posttraumatic osteoarticular infections caused by *Clostridium* spp. are rare, and their outcomes are often unfavorable because of the persistence of the bacteria in bone (1,2). In a recent series of 12 patients (2), only 1 case of posttraumatic osteoarticular infection was caused by *C. tetani* (fracture of the distal humerus with polymicrobial infection). However, no information was available about the production of tetanospasmin by the infecting strain.

¹These authors contributed equally to this article.